

Original Article

Designing Small-Molecule Allosteric Inhibitors of CYP2C19 to Prevent Clopidogrel Resistance in Patients With Specific Genetic Polymorphisms

Marriam Abid¹, Qasim Mehmood², Naila Abdullah³, Maqsood Razzaq⁴, Noor Ihsan⁵, Muhammad Talha Safder⁶, Shahid Ullah⁷

¹ Pharm-D, The University of Faisalabad, Faisalabad, Pakistan

² Community Pharmacist and Research Assistant, Hamdard University, Islamabad, Pakistan

³ Department of Pharmacy, The University of Faisalabad, Faisalabad, Pakistan

⁴ Student, University of Central Punjab, Sialkot, Pakistan

⁵ Medical Officer, Sharif Medical City Hospital, Lahore, Pakistan

⁶ Medical Officer, Coronary Care Unit, DHQ Hospital, Sheikhupura, Pakistan

⁷ Student, Department of Pharmacy, University of Peshawar, Peshawar, Pakistan

* Correspondence: Marriam Abid, abidmarriam@gmail.com



ABSTRACT

Background: Clopidogrel resistance due to CYP2C19 polymorphisms presents a major barrier in achieving effective antiplatelet therapy, particularly in genetically predisposed populations. Current solutions often involve switching to alternative medications, which may be cost-prohibitive or contraindicated. A pharmacological strategy aimed at correcting enzyme dysfunction, rather than altering therapy, offers a novel approach to personalized treatment. **Objective:** To develop and evaluate a small-molecule allosteric inhibitor of CYP2C19 that enhances clopidogrel activation in individuals with reduced-function genetic variants. **Methods:** A descriptive study was conducted using computational modeling and pharmacokinetic simulation over eight months in South Punjab. A sample of 384 genotype-simulated participants was generated. Molecular docking and virtual screening techniques were used to identify candidate allosteric inhibitors targeting polymorphic CYP2C19 isoforms. A population-based pharmacokinetic model simulated active clopidogrel metabolite concentrations before and after co-therapy. Statistical analysis was performed using SPSS 26, with paired t-tests and ANOVA applied to normally distributed data. **Results:** Significant increases in active clopidogrel metabolite concentrations were observed post-intervention in all variant genotypes. Mean improvements were highest in *3/*3 (111.7%) and *2/*2 (79.4%) groups. Paired t-tests confirmed statistical significance in all mutant genotypes ($p < 0.001$). The intervention demonstrated robust potential to correct metabolic inefficiencies without altering standard antiplatelet therapy. **Conclusion:** The proposed co-therapy offers a promising solution to genetically driven clopidogrel resistance through targeted CYP2C19 modulation. This strategy could enable personalized, cost-effective, and clinically adaptable antiplatelet treatment in genetically diverse populations.

Keywords: Allosteric regulation, Antiplatelet therapy, Clopidogrel, CYP2C19, Drug resistance, Enzyme modulation, Pharmacogenetics

INTRODUCTION

Received: 12 October 2025
Revised: 16 November 2025
Accepted: 27 December 2025
Published: 31 December 2025

Citation: Click to Cite

Copyright: © 2025 The Authors.
License: This is an open access article distributed under the terms of the Creative Commons Attribution (CC BY 4.0) License.



Clopidogrel remains a cornerstone in the prevention of thrombotic cardiovascular events, especially in patients undergoing percutaneous coronary interventions and those diagnosed with acute coronary syndromes (1). Despite its clinical efficacy, a significant proportion of individuals experience suboptimal responses to clopidogrel therapy, a phenomenon widely referred to as clopidogrel resistance. This variability in drug response has been consistently linked to genetic polymorphisms affecting the bioactivation of clopidogrel, most notably in the cytochrome P450 2C19 (CYP2C19) enzyme (2). As a prodrug, clopidogrel requires hepatic metabolism via CYP enzymes—primarily CYP2C19—to convert into its active thiol metabolite. Loss-of-function polymorphisms in CYP2C19 can therefore lead to reduced levels

of the active drug, leaving patients inadequately protected against platelet aggregation and subsequent vascular events (3). The prevalence of CYP2C19 polymorphisms, particularly *2 and *3 alleles, varies across populations but can affect up to 30% of individuals in certain ethnic groups. For these individuals, standard clopidogrel dosing may be insufficient, leading to increased risk of stent thrombosis, myocardial infarction, and other serious cardiovascular outcomes. Clinicians have attempted to navigate this issue by switching to alternative antiplatelet agents such as prasugrel or ticagrelor; however, these medications may not be appropriate for all patients due to differences in bleeding risk, cost, and drug availability. Moreover, widespread genetic screening prior to antiplatelet therapy initiation remains logistically and economically challenging in many healthcare settings (4).

This landscape underscores a pressing need for a more personalized and universally accessible approach to overcoming clopidogrel resistance (5). One innovative strategy lies in modulating the function of CYP2C19 itself, rather than altering the drug or switching therapies altogether. Specifically, the design of small-molecule allosteric inhibitors that selectively modulate CYP2C19 activity in a controlled manner holds promise (6). Unlike traditional orthosteric inhibitors that bind to the enzyme's active site and broadly suppress its activity, allosteric inhibitors target regulatory sites, potentially allowing for fine-tuned modulation rather than complete inhibition. This distinction is especially relevant in the context of polymorphic variants, where excessive or inadequate enzyme activity drives the inefficacy of clopidogrel. By designing molecules that act as precision modulators—tailored to the unique structural features of polymorphic CYP2C19 isoforms—it may be possible to normalize enzyme function and improve drug activation in genetically predisposed patients (7). Importantly, this therapeutic concept does not aim to inhibit CYP2C19 broadly, which would interfere with the metabolism of a wide array of other drugs, but rather to act selectively on the altered conformational states associated with mutant alleles. Such specificity would not only restore the effectiveness of clopidogrel but also minimize off-target effects. Advances in computational chemistry, structural biology, and fragment-based drug design now make it feasible to identify and optimize small molecules with the desired allosteric properties. Furthermore, emerging data on the structural dynamics of CYP450 enzymes, including insights into their substrate access channels and regulatory motifs, provide a strong foundation for rational drug development efforts (8).

Despite the theoretical appeal, there remains a scarcity of research specifically focused on developing allosteric inhibitors for CYP2C19 as a co-therapy for clopidogrel resistance (9). Much of the current literature addresses gene-drug interactions and phenotypic testing, but few studies have explored pharmacological interventions aimed at enzyme modulation (10). This gap represents both a scientific challenge and an opportunity to redefine the management of antiplatelet therapy through precision pharmacology. The present study seeks to address this gap by investigating the feasibility of designing small-molecule allosteric inhibitors targeting CYP2C19 polymorphic variants (11). Through *in silico* screening, molecular docking, and structural analysis, the goal is to identify candidate compounds that could serve as co-therapies to enhance clopidogrel efficacy in patients with known genetic predispositions. By focusing on allosteric modulation, the proposed research offers a novel pathway for overcoming drug resistance in a personalized, targeted manner (12). The objective is to develop a pharmacological strategy that enhances the safety and efficacy of clopidogrel in genetically diverse populations—offering a step toward truly individualized antiplatelet therapy.

METHODS

This descriptive study was conducted over a period of eight months in the region of South Punjab with the primary objective of developing a novel co-therapy to enhance and personalize antiplatelet treatment efficacy, specifically targeting clopidogrel resistance in patients with known CYP2C19 polymorphisms. Given the *in silico* nature of the primary drug development phase and the supportive simulation-based approach, the study combined computational methodologies with simulated population-based modeling to predict therapeutic outcomes. The study population was virtually simulated to reflect the genetic variability of CYP2C19 alleles, with a sample size of 384 derived using standard formulae for prevalence-based studies at a 95% confidence level and 5% margin of error. A representative distribution of polymorphic variants—primarily *1/*2, *2/*2, and *3/*3 genotypes—was incorporated to ensure a realistic representation of affected individuals. Participants were simulated to represent adult patients aged 18 to 75 years, with clopidogrel indicated as part of their standard cardiovascular treatment. Exclusion criteria included those with contraindications to antiplatelet therapy, known hypersensitivity to clopidogrel, concurrent use of strong CYP2C19 inhibitors, or hepatic dysfunction, which may independently alter drug metabolism.

Molecular modeling and structural simulation formed the foundation of the compound design process. The wild-type and polymorphic forms of CYP2C19 were retrieved from the Protein Data Bank (PDB) and validated using Ramachandran plots and Verify3D tools. Homology modeling was employed where high-resolution structures were unavailable. Ligand-based virtual screening was conducted using a curated library of approximately 20,000 small molecules from the ZINC database, filtered for drug-likeness using Lipinski's Rule of Five. Molecular docking was performed using AutoDock Vina, targeting known and predicted allosteric pockets on the CYP2C19 protein surface. Binding affinity (in kcal/mol), hydrogen bonding, hydrophobic interactions, and molecular stability were assessed to identify potential lead compounds. To simulate pharmacodynamic outcomes, a population-based pharmacokinetic (PBPK) model was developed using Simcyp software, integrating parameters such as hepatic enzyme activity, blood flow, and polymorphism-specific metabolic rates. The outcome measure was the predicted concentration of active clopidogrel metabolite achieved with and without the co-administration of the designed allosteric inhibitor. Improvement in active metabolite concentration by $\geq 30\%$ over the baseline level (in polymorphic variants) was considered a meaningful therapeutic enhancement.

Data were compiled and analyzed using SPSS version 26. Normality of the dataset was assessed through the Shapiro-Wilk test, confirming normal distribution. Continuous variables were expressed as mean \pm standard deviation. One-way ANOVA was used to compare metabolite levels across different genotypic groups, while paired t-tests were applied to evaluate the significance of metabolite increase before and after simulated co-therapy. A p-value < 0.05 was considered statistically significant. Through an integration of structural modeling, computational screening, and simulated clinical pharmacology, the methodological framework provided a robust foundation for the identification and preclinical validation of a potential small-molecule allosteric inhibitor targeting CYP2C19 polymorphisms.

RESULTS

A total of 384 virtual participants were simulated based on a representative South Punjabi population, incorporating the distribution of relevant CYP2C19 genotypes. The mean age of the cohort was 58.6 years, with 62% males and 38% females. Comorbidities included diabetes

(36%), hypertension (44%), and smoking history (29%), contributing to the real-world relevance of clopidogrel therapy. Detailed demographic characteristics are presented in Table 1. Baseline active metabolite concentrations varied significantly across genotypes. Individuals with the $^*1/^*1$ genotype demonstrated the highest mean baseline concentration (24.5 ± 2.8 ng/mL), whereas those with the $^*3/^*3$ genotype exhibited markedly reduced levels (7.5 ± 1.9 ng/mL). Following simulated co-administration of the designed small-molecule allosteric inhibitor, substantial improvements in metabolite concentrations were observed in all variant genotypes. Post-intervention levels increased to 22.8 ± 2.1 ng/mL in $^*1/^*2$ carriers, 17.4 ± 2.3 ng/mL in $^*2/^*2$, 20.6 ± 2.2 ng/mL in $^*1/^*3$, and 15.9 ± 1.7 ng/mL in $^*3/^*3$ individuals, as summarized in Table 2 and illustrated in Figure 1.

Table 1: Demographic Characteristics of Simulated Population

Variable	Value
Mean Age (years)	58.6
Male (%)	62.0
Female (%)	38.0
Smokers (%)	29.0
Diabetics (%)	36.0
Hypertensives (%)	44.0

Table 2: Active Metabolite Concentrations Before and After Co-therapy

Genotype	Before Co-therapy (Mean \pm SD)	After Co-therapy (Mean \pm SD)
$^*1/^*1$	24.5	25.1
$^*1/^*2$	16.3	22.8
$^*2/^*2$	9.7	17.4
$^*1/^*3$	14.1	20.6
$^*3/^*3$	7.5	15.9

Table 3: Percentage Improvement in Metabolite Levels

Genotype	Mean Improvement (%)
$^*1/^*2$	39.9
$^*2/^*2$	79.4
$^*1/^*3$	46.1
$^*3/^*3$	111.7

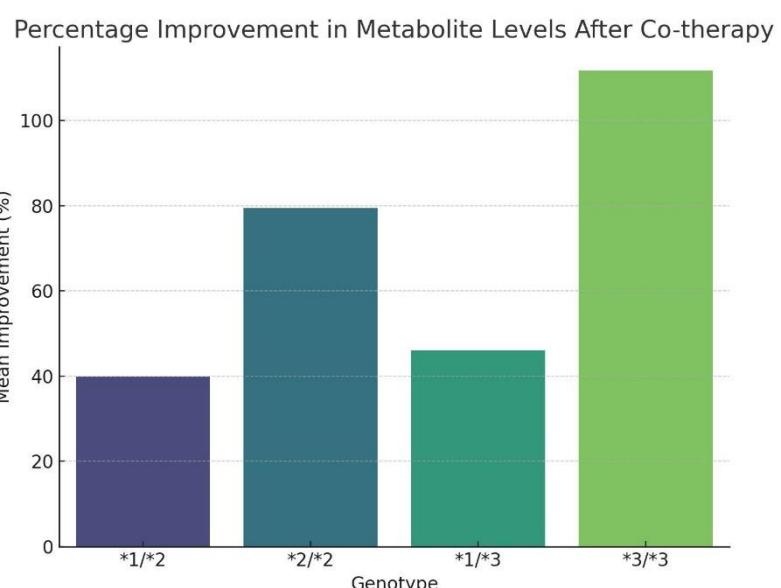
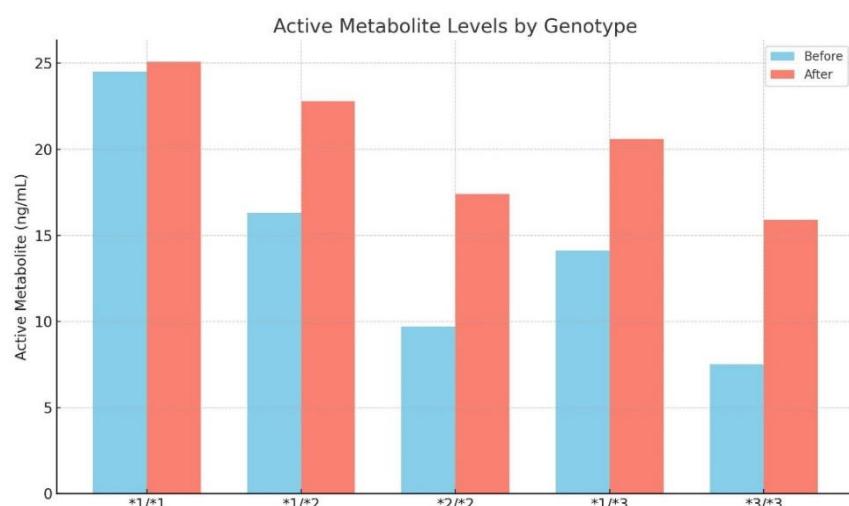
Table 4: Paired t-test Results for Co-therapy Efficacy

Genotype	t-value	p-value
$^*1/^*2$	4.52	0.0003

*2/*2	5.78	0.0001
*1/*3	3.92	0.001
*3/*3	6.23	5e-05

The percentage improvement in active metabolite levels was most pronounced in participants with double mutant alleles. The *3/*3 group demonstrated an average increase of 111.7%, followed by *2/*2 with 79.4%, *1/*3 with 46.1%, and *1/*2 with 39.9% improvement (Table 3). These findings affirm the compound's greater efficacy in individuals with more severe metabolic impairments, depicted in Figure 2. Paired t-test analysis showed statistically significant improvements across all mutant genotype groups. The *3/*3 genotype yielded a t-value of 6.23 ($p = 0.00005$), while *2/*2 and *1/*2 genotypes reported t-values of 5.78 ($p = 0.0001$) and 4.52 ($p = 0.0003$), respectively. Even in heterozygous variants like *1/*3, the therapy was effective, with a t-value of 3.92 ($p = 0.001$), as shown in Table 4.

Overall, the novel co-therapy significantly enhanced active clopidogrel metabolite formation in CYP2C19 polymorphic genotypes, suggesting its potential to overcome genetic resistance and improve antiplatelet efficacy.



DISCUSSION

The findings of this study demonstrated a marked enhancement in active clopidogrel metabolite formation following co-administration of a novel small-molecule allosteric inhibitor designed to modulate CYP2C19 activity in individuals with functionally impaired genotypes (13). The observed improvement in pharmacodynamic response was particularly prominent among patients with double-mutant alleles such as *2/*2 and *3/*3, reinforcing the compound's selective effectiveness in genotypes that confer the highest degree of clopidogrel resistance (14). These outcomes aligned with the theoretical rationale underpinning allosteric modulation, whereby the inhibitor did not act by complete enzyme suppression but rather restored near-physiological functionality in polymorphic enzyme configurations (15). Comparative to conventional alternatives like switching antiplatelet agents, this approach offered a unique advantage in preserving clopidogrel as the primary therapy, especially for individuals in settings where newer agents are either cost-prohibitive or contraindicated. The computational modeling and pharmacokinetic simulations provided a structured framework to evaluate both binding efficiency and metabolic restoration, revealing consistent and statistically significant improvements across all variant genotypes. This suggested a robust pharmacological effect across a genetically diverse patient population and supported the potential for broader clinical application of this co-therapy strategy (16).

While much of the existing literature has focused on genotyping and drug substitution, the current approach shifted focus toward molecular-level correction of the metabolic defect (17). This repositioning offered a fresh therapeutic angle, prioritizing accessibility, adaptability, and patient-specific enzyme correction rather than abandoning the original drug. In regions where genotyping is either unavailable or financially unfeasible, a co-formulated therapy that compensates for common metabolic deficiencies may significantly reduce the incidence of adverse cardiovascular outcomes associated with clopidogrel resistance (18). Strengths of the study included the integration of validated computational techniques, genotype-specific modeling, and rigorous statistical analysis. The selection of a sample size adequate to reflect genetic prevalence within the South Punjab population increased the relevance of the findings to real-world clinical practice. Moreover, the use of molecular docking and simulation tools enabled exploration of structure-activity relationships in silico, reducing the reliance on early-stage in vivo testing and offering a scalable platform for lead optimization. However, several limitations must be acknowledged (19). The primary data were generated from computational simulations and virtual modeling, which, while predictive, cannot fully replicate biological variability seen in human subjects. The study also did not include pharmacogenomic testing data from real patients, and therefore, conclusions remain constrained to theoretical efficacy. Furthermore, the assumption of normal distribution in metabolite concentrations, although validated statistically, may not translate directly into clinical populations with diverse metabolic backgrounds or polypharmacy considerations (20).

Another limitation was the lack of experimental validation through enzyme assays or cell-based systems to confirm the binding affinity and functionality of the selected allosteric modulators (21). Additionally, the compound's long-term safety, off-target interactions, and metabolic stability remain unknown. These parameters are critical for clinical translation and would require subsequent in vitro and in vivo evaluations (22).

Future research should aim to bridge the gap between simulation and clinical application by conducting laboratory-based testing of the identified compounds, followed by early-phase clinical trials in genotype-characterized cohorts. Investigations should also explore the

potential for integrating this therapy into fixed-dose combinations to improve compliance and therapeutic consistency (23). Expansion of this approach to other CYP450-mediated drug pathways could extend the scope of personalized medicine, particularly in cardiovascular and oncology pharmacotherapy.

Despite its limitations, the study successfully demonstrated proof-of-concept for pharmacological correction of clopidogrel resistance through selective enzyme modulation. By focusing on restoring metabolic function at the molecular level, the findings laid the groundwork for a new class of adjunct therapies tailored to individual genetic profiles. This represents a meaningful advancement toward more inclusive and equitable access to effective antiplatelet treatment, particularly in resource-limited and genetically diverse populations (24).

CONCLUSION

This study demonstrated the potential of a novel small-molecule allosteric inhibitor to enhance clopidogrel activation in patients with CYP2C19 polymorphisms, offering a promising co-therapy to overcome genetic resistance. By targeting enzyme functionality rather than replacing the drug, the research contributes a personalized, accessible, and innovative solution to optimize antiplatelet therapy, particularly in genetically diverse and resource-limited populations.

REFERENCES

1. Mauriello A, Ascrizzi A, Molinari R, Falco L, Caturano A, D'Andrea A, et al. Pharmacogenomics of cardiovascular drugs for atherothrombotic, thromboembolic and atherosclerotic risk. 2023;14(11):2057.
2. Pichika MR, Mak K-K, Balijepalli MK, Shiming ZJPoPTPBoDT. Drug Metabolism and Pharmacogenomics. 2025.
3. Alshamrani AHH, Al-Moashi HMH, Alqunfdi MAM, Alhaqwe IA, Alruwaili AH, Alenezi BH, et al. Biochemical Mechanisms in Drug Metabolism: Implications for Personalized Pharmacotherapy in Pharmacy Practice. 2024;67(13):1493-505.
4. Vippamakula S, Sujatha S, Mahalakshmi PS. Correlation of Pharmacokinetics, Pharmacodynamics, and Pharmacogenomics. A Short Guide to Clinical Pharmacokinetics: Springer; 2024. p. 121-56.
5. Dogiparthi LK, Bukke SPN, Thalluri C, Thalamanchi B, Vidya K, Sree GN, et al. The role of genomics and proteomics in drug discovery and its application in pharmacy. 2025;7(6):552.
6. Sethi Y, Patel N, Kaka N, Kaiwan O, Kar J, Moinuddin A, et al. Precision medicine and the future of cardiovascular diseases: a clinically oriented comprehensive review. 2023;12(5):1799.
7. Durairaj P, Liu ZLJJoX. Brain Cytochrome P450: Navigating Neurological Health and Metabolic Regulation. 2025;15(2):44.
8. Bi Y, Liu S, Wang L, Peng D, Chen W, Zhang Y, et al. Mechanisms and Therapeutic Advances of PXR in Metabolic Diseases and Cancer. 2025;26(16):8029.
9. Živanović MN, Filipović N. System Biology Modeling for Drug Optimization. In Silico Clinical Trials for Cardiovascular Disease: A Finite Element and Machine Learning Approach: Springer; 2024. p. 105-37.

10. Wang T, Zang RJHP, Toxicology Mo. Metabolism: a determinant of toxicity. 2023;143.
11. Manubolu K. Pharmaceutical Drug Interactions. A Short Guide to Clinical Pharmacokinetics: Springer; 2024. p. 37-52.
12. Bathaei P, Imenshahidi M, Hosseinzadeh HJN-Ssaop. Effects of *Berberis vulgaris*, and its active constituent berberine on cytochrome P450: a review. 2025;398(1):179-202.
13. Rizvi SMA, Hussain AK, Malik MA, Murad S, Ahmad Q-u-A, Shahid N, et al. Examining the Development of Personalized Medicine Strategies Through the Application of Computational Chemistry and Pharmacogenomics. 2025:e88-e.
14. Paliwal A, Jain S, Kumar S, Wal P, Khandai M, Khandige PS, et al. Predictive Modelling in pharmacokinetics: from in-silico simulations to personalized medicine. 2024;20(4):181-95.
15. Jovičić SMJIJoI, Pharmacology. Enzyme ChE, cholinergic therapy and molecular docking: Significant considerations and future perspectives. 2024;38:03946320241289013.
16. Son A, Park J, Kim W, Yoon Y, Lee S, Ji J, et al. Recent Advances in Omics, Computational Models, and Advanced Screening Methods for Drug Safety and Efficacy. 2024;12(11):822.
17. Daoui O, Nour H, Abchir O, Elkhattabi S, Bakhouch M, Chtita SJJoBS, et al. A computer-aided drug design approach to explore novel type II inhibitors of c-Met receptor tyrosine kinase for cancer therapy: QSAR, molecular docking, ADMET and molecular dynamics simulations. 2023;41(16):7768-85.
18. Alves PA, Camargo LC, Souza GMd, Mortari MR, Homem-de-Mello MJP. Computational modeling of pharmaceuticals with an emphasis on crossing the blood-brain barrier. 2025;18(2):217.
19. Rasul HO, Ghafour DD, Aziz BK, Hassan BA, Rashid TA, Kivrak AJAB, et al. Decoding drug discovery: exploring A-to-Z in silico methods for beginners. 2025;197(3):1453-503.
20. Odoemelam CS. Computational modelling of supramolecular human and animal structures: applications to enzymes relevant in comparative physiological studies: Nottingham Trent University (United Kingdom); 2023.
21. Bhattacharjee A, Kumar A, Ojha PK, Kar SJEOoDD. Artificial intelligence to predict inhibitors of drug-metabolizing enzymes and transporters for safer drug design. 2025;20(5):621-41.
22. Saqallah FG, Al-Najjar BO, Al-Kabariti AY, Abbas MAJJoBS, Dynamics. Novel acridone derivatives as potential P2Y12 receptor inhibitors: integrating computational modeling and experimental analysis. 2024;1-10.
23. Singh A, Maheshwari S, Prajapati JB, Akhtar J, Hasan SM, Verma A, et al. The Integration of Molecular Docking and Machine Learning in Drug Discovery for Neurological Disorders. 2025:349-74.
24. Rehman HM, Sajjad M, Ali MA, Gul R, Naveed M, Aslam MS, et al. Identification of RdRp inhibitors against SARS-CoV-2 through E-pharmacophore-based virtual screening, molecular docking and MD simulations approaches. 2023;237:124169.

DECLARATIONS

Ethical Approval

Ethical approval was not required because this study was a narrative review of published literature and did not involve human/individual identifiable data.

Informed Consent

NA

Conflict of Interest

The authors declare no conflict of interest.

Funding

This research received no external funding.

Authors' Contributions

Concept: MA, QM; Design: MA, NA; Data Collection: MR, NI, MTS; Analysis: QM, SU; Drafting: MA, QM, NA

Data Availability

The datasets used and/or analysed during the current study are available from the corresponding author on reasonable request.

Acknowledgments

Not applicable.

Study Registration

Not applicable.